

0960-894X(94)00164-2

## DESIGN AND SYNTHESIS OF A PROTOTYPE MODEL ANTAGONIST OF TACHYKININ NK-2 RECEPTOR

Stephen Hanessian\* and Baptiste Ronan†
Department of Chemistry, Université de Montréal
P.O. Box 6128, Succ. Centre-ville, Montréal, P.Q., CANADA, H3C 3J7

## Abdel Laoui

Rhône-Poulenc Rorer S.A., Central Research, Centre de Recherche de Vitry-Alfortville, 13, quai Jules Guesde - BP 14, 94403 Vitry-sur-Seine Cedex FRANCE

**Abstract:** A conformationally biased bicyclic lactam containing strategically placed hydrophobic groups was synthesized as a prototype model antagonist molecule of the NK-2 receptor.

Tachykinins, including the undecapeptide substance P and related neurokinins, are generally associated with physiological events leading to pain and inflammation.<sup>1,2</sup> These mediators of neurotransmission or neuromodulation in the central and peripheral nervous systems have affinities for certain receptors designated as NK-1, 2 and 3.<sup>3</sup> Substance P and the decapeptides, neurokinin A and B have a common C-terminal sequence consisting of Phe-X-Gly-Leu-MetNH<sub>2</sub>.<sup>4</sup> This minimal structural information, in conjunction with extensive studies on peptide and non-peptide analogs,<sup>5</sup> have led to the development of antagonists<sup>6</sup> to these tachykinins particularly those with high affinities to NK-1 and NK-3 receptors.<sup>7</sup> Although somewhat more elusive, selective antagonists to neurokinin A at the NK-2 receptor have also been reported.<sup>8</sup> A recent conformational study<sup>9</sup> in solution of a cyclic hexapeptide potent antagonist by NMR techniques has revealed information regarding the shape and relative disposition of the pendant groups as well as the existence of a β-turn subunit involving Trp-Phe-Gly-Leu residues (Figure 1).

Based on these observations, and data available from the literature on some of the more potent and selective antagonists of NK-2,5-8 we have derived a prototype for an antagonist model which is shown in expression 1 (Figure 1). The intention was to deploy hydrophobic groups that could simulate the Trp-Phe portion, on a rigid

Figure 1

<sup>†</sup> Present address, Rhône-Poulenc Rorer S.A., Central Research, Centre de Recherche de Vitry-Alfortville, 13, quai Jules Guesde - BP 14, 94403 Vitry-sur-Seine Cedex FRANCE

a) H<sub>2</sub> - 4 Atm, Pt/C (5%), Toluene, rt, 110h; cis:trans = 9:1, separation by flash chromatography on silica gel (AcOEt/hexane 5/1). b) Boc<sub>2</sub>O (1.1 eq), (iPr<sub>2</sub>)NEt (1.1 eq), CH<sub>2</sub>Cl<sub>2</sub>, rt, 18h, 50% (2 steps). c) H<sub>2</sub> - 1 Atm, Pd/C (10%), MeOH, rt, 1h. d) N-Methyl morpholine (1 eq), isobutyl chloroformate (1 eq), THF, -20°C, 20 min; 2-mercaptopyridine N-oxide, sodium salt hydrate (1.1 eq), -20°C in the dark 1h (quant.). e) Methyl acrylate (5 eq) hv 250 W, rt, 30 min, 80%. f) TFA (20 eq), CH<sub>2</sub>Cl<sub>2</sub>, rt, 16h; NH<sub>4</sub>OH. g) Me<sub>3</sub>Al (2 eq), CH<sub>2</sub>Cl<sub>2</sub>, rt, 1h, 50% (2 steps). h) mCPBA (1 eq), CHCl<sub>3</sub>, 0°C, 1h; toluene, 110°C, 1h, 82%. i) benzyl phenyl sulfide (1.1 eq), HMPA (2.5 eq), nBuLi (1.1 eq), THF, -78°C, 1h, -78°C, 1h, -78°C, 69%. j) W<sub>2</sub> Raney-Ni, EtOH, rt, 1h, 95%. k) PhCH<sub>2</sub>NHAlMe<sub>2</sub> (3 eq), CH<sub>2</sub>Cl<sub>2</sub>, rt, 18h; aq. HCl, 50%. l) LDA (2 eq), THF, -10°C, 30 min; trisyl N<sub>3</sub> (2 eq), THF, -78°C, 5 min; AcOH (10 eq), -78°C; 30°C, 2h; separation by flash chromatography on silica gel (AcOEt/cyclohexane=1:1), 50%. m) H<sub>2</sub> - 4 Atm, Pd/C (10%), rt, 24h. n) indole -3-acetic acid (1.2 eq), EDCI (1.2 eq), DMF, rt, 16h, 36%.

bicyclic template that would act as a surrogate for a β-turn<sup>10</sup> much same as in the cyclic hexapeptide.<sup>9</sup> The C-terminal benzylamide group can also be found in a number of NK-1 and NK-2 selective peptidic antagonists,<sup>7,8</sup>

In order to secure a high level of internal asymmetric induction by a resident group, we chose Lpyroglutamic acid 2 as a logical starting material (Scheme 1). A sequence of known reactions 11 led to the diester 3 in good overall yield. Catalytic hydrogenation gave a 9:1 cis/trans mixture of the expected pyrrolidine derivatives which were separated by chromatography.<sup>12</sup> Protection of the amine as the Boc derivative, and catalytic debenzylation of the benzyl ester gave a monoester which was transformed into the corresponding thiopyridinyl hydroxamate ester 5. Application of the Barton procedure 13 using methyl acrylate as a radical acceptor led to 6 in excellent overall yield. Unfortunately, the reaction failed when crotonate or 3-benzyl acrylate esters were utilized as radical Michael acceptors, in accord with previous observations. 14,15 The Barton reaction allowed the introduction of the required three-carbon acid branch with a functionally useful pyridylthio group for subsequent manipulation. Thus, removal of the N-Boc group and cyclization afforded the bicyclic lactam 7 as a mixture of two diastereomers. Oxidative elimination 15,16 led to the unsaturated lactam 8. In view of the reluctance of cuprate-type reagents to cleanly add to the  $\alpha,\beta$ -unsaturated lactam 8, we resorted to a two-step, albeit, efficient protocol for the introduction of the C-benzyl group. Thus, treatment of 8 with benzylphenylsulfide anion in THF-HMPA<sup>17</sup> led to 9 as a mixture of diastereomers in good yield and excellent selectivity based on the NMR analysis of subsequent intermediates. In the absence of HMPA, a significant quantity of addition to the ester function occurred also. Removal of the phenylthio group by desulfurization gave 10 as an oil. There now remained to introduce the amino group in the lactam subunit and to complete the synthesis of our intended target molecule. As an obvious choice, we turned to the electrophilic transfer of azide from an arylsulfonyl azide 18-20 to the enolate generated from 10. The latter proved somewhat sluggish to form under a variety of conditions. In fact treatment of 10 with LiHMDS in THF with trisyl or tosyl azide at -78°C, led to the introduction of an azide group adjacent to the ester function rather than at the desired site in 42% yield. Introduction of the benzylamide group using the Weinreb protocol<sup>21</sup> gave 11, as a crystalline solid. Electrophilic addition of azide ion to the dianion generated from 11 at the desired site was now possible, resulting in a 1:1 mixture of 12 and 13. Various attempts to change this ratio were not successful (solvent, temperature, azide source). The moderate yields in such reactions with dianions have been previously reported.<sup>22,23</sup> Although the two isomers were separable by chromatography, it was not possible to efficiently epimerize the undesired isomer 12 into 13. Catalytic reduction of the trans isomer 13, followed by acylation gave the intended target molecule 1 as a crystalline solid.<sup>24</sup> Although the dimensions of the crystals were not suitable for single crystal X-ray analysis, the structure of 1 was unambiguously assigned from detailed NMR analysis.<sup>24</sup> Furthermore, the presence of intramolecular H-bonding was strongly suggested from the FT-IR spectrum of 1 (3309 cm<sup>-1</sup>, 3400 cm<sup>-1</sup>).<sup>25</sup>

The prototype model antagonist 1 had no affinity for NK-1 receptor, and showed a low affinity for human recombinant NK-2 receptor with an affinity binding of  $3\mu M$ ,  $^{26}$  which is a promising beginning for a new "designed" structure. We believe that further fine-tuning of the proposed basic template structure 1 may be possible, by attachment of other pendant groups on rings A and B, coupled with new information from the structures of emerging antagonists. Hopefully this will eventually lead to a new generation of "small", potent, and selective antagonists of these fascinating receptors.  $^{27}$ 

Acknowledgement. We thank NSERCC, FCAR and Rhône-Poulenc Rorer (Vitry, France) for financial assistance. We acknowledge useful discussions with Drs. J.-L. Malleron and we thank Dr. V. Fardin for the receptor binding assays.

## References and Notes

1. For selected reviews, see Reese, D.C. Ann. Rep. Med. Chem., 1993, 28, 59; Maggi, A.-C.; Quartara, L.; Giuliani, S.; Patacchini, R. Drugs of the Future, 1993, 18, 155; Watling, K.J.; Guard, S. Neurotransmissions, 1992, 26, 43.

- Lowe III, J.A.; Snider, R.M. Ann. Rep. Med. Chem., 1993, 28, 99.
   Hopkins, B.; Powell, S.J.; Danks, P.; Briggs, I.; Graham, A. Biochem. Biophys. Res. Commun., 1991, 180, 1110; Takeda, Y.; Chou, K.B.; Takeda, J.; Sachais, S.; Krause, J.E. Biochem. Biophys. Res. Commun., 1991, 179, 1232;
- Shigemoto, R.; Chou, K.B.; Fakeda, J.; Sachais, S.; Krause, J.E. Biochem. Biophys. Res. Commun., 1991, 179, 1232;
   Shigemoto, R.; Yokota, Y.; Tsuchida, K.; Nakanishi, S. J. Biol. Chem., 1990, 265, 623; and references cited therein.
   Chang, M.M.; Leeman, S.E.; Niall, H.D. Nature, 1971, 232, 86.
   For recent reviews on peptidomimetics see, Giannis, A.; Kolter, T. Angew. Chem., Int. Ed. Engl., 1993, 32, 1244;
   Kahn, M. Synlett, 1993, 821; Olson, G.L. et al. J. Med. Chem., 1993, 36, 3039; Hirschmann, R. Angew. Chem. Ed. Engl. 1991, 30, 1278; Hölzemann, G. Kontakte, 1991, 3; Ball, J.B.; Alewood, P.F. J. Mol. Recogn., 1990, 3, 55; Morgan, B.A.; Gainor, J.A. Ann. Rep. Med. Chem., 1989, 24, 243; Kahn, M.; Wilke, S.; Chen, B.; Fujita, K.; Lee, Y.-H.; Johnson, M.E. J. Mol. Recogn., 1988, 1, 75.
   McFlroy, A. B. Current Drugs, 1993, 1157.
- McElroy, A.B. Current Drugs, 1993, 1157.
- McElroy, A.B. Current Drugs, 1993, 1157.
   For selected recent examples, see Hagiwara, D.; Miyake, H.; Morimoto, H.; Murai, M.; Fujii, T.; Matsuo, M. J. Med. Chem., 1992, 35, 2015; Ward, P.; Ewan, G.B.; Jordan, C.C.; Ireland, S.J.; Hagan, R.M.; Brown, J.R. J. Med. Chem., 1990, 33, 1848; Folkers, K.; Feng, D.M.; Asano, N.; Hakanson, R.; Weisenfeld-Hallin, Z.; Leander, S.; Proc. Natl. Acad. Sci. USA, 1990, 87, 4833.
   For selected examples, see, Williams, B.J.; Curtis, N.R.; McKnight, A.T.; Maguire, J.J.; Young, S.C.; Veber, D.F.; Baker, R. J. Med. Chem., 1993, 36, 2; Hale, J.J.; Finke, P.E.; MacCoss, M. Bioorg. Med. Chem. Letters, 1993, 3, 319; Harbeson, S.L.; Shatzer, S.A.; Le, T.-B.; Buck, S.H. J. Med. Chem., 1992, 35, 3949; McElroy, A.B.; Clegg, S.P.; Deal, M.J.; Ewan, G.B.; Hagan, R.M.; Ireland, S.J.; Jordan, C.C.; Porter, B.; Ross, B.C.; Ward, P.; Whittington, A.R. J. Med. Chem., 1992, 35, 2582; Advenier, C.; Rouissi, N.; Nguyen, Q.T.; Emonds, Alt, X.; Breliere, J.-C.; Neliat, G.; Naline, E.; Regoli, D. Biochem. Biophys. Res. Commun., 1992, 184, 1418; Morimoto, H.; Murai, M.; Maeda, Y.; Yamaoka, M.; Nishikawa, M.; Kiyotoh, S.; Fujii, T. J. Pharmacol. Exptl. Therap. 1992, 282, 398; Rovero, P.; Astolfi, M.; Renzetti, A.R.; Patacchini, R.; Giachetti, A.: Maggi, C.A. Peptides, 1991, 12, 1015.
   Malikayil, J.A.; Harbeson, S.L. Int. J. Peptide Protein Res., 1992, 39, 497.
- 9. Malikayil, J.A.; Harbeson, S.L. Int. J. Peptide Protein Res., 1992, 39, 497.

  10. See for example Nagai, U.; Sato, K. Tetrahedron Lett., 1985, 26, 647; Moeller, K.D.; Hanau, C.E.; d'Avignon, A.
- Tetrahedron Lett., 1994, 35, 825.

  11. Fang, F.G.; Danishefsky, S.J. Tetrahedron Lett., 1989, 30, 3621; Nagasaka, T.; Tsukada, A.; Hamaguchi, F. Heterocycles, 1986, 24, 2015.
- 12. For a related reduction, see Petersen, J.S.; Fels, G.; Rapoport, H. J. Am. Chem. Soc., 1984, 106, 4539.
- For a related reduction, see Petersen, J.S.; Fels, G.; Rapoport, H. J. Am. Chem. Soc., 1984, 100, 4539.
   Barton, D.H.R., Hervé, Y.; Potier, P.; Thierry, J. Tetrahedron, 1987, 43, 4297; Barton, D.H.R. Aldrichimica Acta, 1990, 23, 3; Crich, D.; Quintero, L. Chem. Rev., 1989, 89, 143.
   Sumi, K.; DiFabio, R.; Hanessian, S. Tetrahedron Lett., 1992, 33, 749.
   Barton, D.H.R.; Togo, H.; Zard, S.Z. Tetrahedron Lett., 1985, 26, 6349.
   Barton, D.H.R.; Togo, H.; Zard, S.Z. Tetrahedron, 1985, 41, 5507.
   Yanagida, M.; Hashimoto, K.; Ishida, M.; Shinosaki, H.; Shirahama, H. Tetrahedron Lett., 1989, 30, 3799.
   Evans, D.A.; Britton, T.C. J. Am. Chem. Soc., 1987, 109, 6881; Evans, D.A.; Ellman, J.A.; J. Am. Chem. Soc., 1000, 111, 1062

- 1989, 111, 1063.
- 19. Wasserman, H.H.; Hlasta, D.J. J. Am. Chem. Soc., 1978, 100, 6780; Wasserman, H.H.; Hlasta, D.J.; Tremper,
- A.W.; Wu, J.S. J. Org. Chem., 1981, 46, 2999.

  20. Kühlein, K.; Jensen, H. Justus Liebigs Ann. Chem., 1974, 369; see also Nishida, A.; Shibasaki, M.; Ikegami, S. Tetrahedron Lett., 1984, 25, 765; Shibuya, M.; Jonbo, Y., Kubota, S. Chem. Pharm. Bull., 1984, 32, 1303.
- Basha, A.; Lipton, M.; Weinreb, S.M. Tetrahedron Lett., 1977, 4171.

- Bevilacqua, P. F.; Keith, D.D.; Roberts, J.L. J. Org. Chem., 1984, 49, 1430.
   Hanessian, S.; Vanasse, B.; Yang, H.; Alpegiani, M. Can. J. Chem., 1993, 71, 1407.
   NMR and spectroscopic data for 1. <sup>1</sup>H NMR d: 8.18 (s, br, 1H), 7.63 (d, 1H, J = 9 Hz), 7.43 (d, 1H, J = 8 Hz), NMR and spectroscopic data for 1. <sup>1</sup>H NMR d: 8.18 (s, br, 1H), 7.63 (d, 1H, J = 9 Hz), 7.43 (d, 1H, J = 8 Hz), 7.32-7.12 (m, 12H), 6.96 (d, 2H, J = 7 Hz), 6.23 (d, 1H, J = 8.6 Hz), 4.49 (t, 1H, J = 9.7 Hz), 4.42 (d, 1H, J = 8 Hz), 4.35 (d, 2H, J = 5.9 Hz), 3.82 (s, 2H), 3.72-3.42 (m, 1H), 2.94-2.88 (dAB, 1H, JAB = 15 Hz, J = 3.8 Hz), 2.54-2.46 (dAB, 1H, JAB = 14 Hz, J = 10 Hz), 2.45-2.39 (m,1H), 2.11-2.05 (m,1H), 1.95-1.40 (m, 5H); <sup>13</sup>C NMR d: 172.39 (CO), 170.40 (CO), 169.81 (CO), 139.07 (C), 138.35 (C), 136.32 (C), 128.92 (CH), 128.40 (CH), 128.32 (CH), 127.31 (CH), 127.10 (CH), 127.07 (C), 126.16 (CH), 123.93 (CH), 122.50 (CH), 120.03 (CH), 118.67 (CH), 111.35 (CH), 108.66 (C), 59.58 (CH), 55.08 (CH), 53.53 (CH), 43.29 (CH<sub>2</sub>), 40.34 (CH), 39.57 (CH<sub>2</sub>), 33.31 (CH<sub>2</sub>), 33.23 (CH<sub>2</sub>), 32.10 (CH<sub>2</sub>), 27.98 (CH<sub>2</sub>); MS (FAB+) m/e 130, 157, 181, 215, 378, 535 (M)
- See for example, Dado, G.P.; Gellman, S.H. J. Am. Chem. Soc., 1993, 115, 4228.
   See for example, Dado, G.P.; Gellman, S.H. J. Am. Chem. Soc., 1993, 115, 4228.
   Fardin, V.; Jolly, A.; Bellot, F.; Crespo, A.; Vaissade, F.; Third Annual Meeting of the European Neuropeptide Club, Cambridge, U.K., April 1993; Neuropeptides, 1993, 24, 232, C21; Guillaume, J.M.; Bellot, F.; Jolly, A.; Coultreault, N.; Fallourd Soria, H.M.; Gravier, I.; Benoist, S.; Fardin, V.; Crespo, A. Baculovirus Expression of United Neuropeptide Club. Cambridge, U.K. April, 1993; Binding Human NK2 Receptor, Third Ann. Meeting Europ. Neuropeptide Club, Cambridge, U.K. April, 1993; Binding assays to human recombinant NK2 receptor was adapted from Bergstrom using <sup>125</sup>I NKA (100 pM) as the radioligand. Samples were incubated at 25°C for 60 min. and the binding interaction was terminated by rapid filtration over glass fiber filters presoaked with 0.3% polyethyleneimine, see Bergstrom, L. et al. *Mol. Pharmacol.*, 1987, 32, 764.
- 27. All new compounds were adequately characterized by spectroscopic and microanalytical methods. Optical rotations were recorded at 25°C.